

# Update on USDA Fertility Project



Megan Rolf  
Oklahoma State University  
2015 NBCEC Brown Bagger

# IDENTIFICATION AND MANAGEMENT OF ALLELES IMPAIRING HEIFER FERTILITY WHILE OPTIMIZING GENETIC GAIN IN ANGUS CATTLE

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USDA-NIFA Award #2013-68004-20364



JF Taylor, DS Brown, MF Smith, RD Schnabel,  
SE Pooch, JE Decker, FD Dailey, and DJ Patterson  
University of Missouri

AL Van Eenennaam  
University of California, Davis

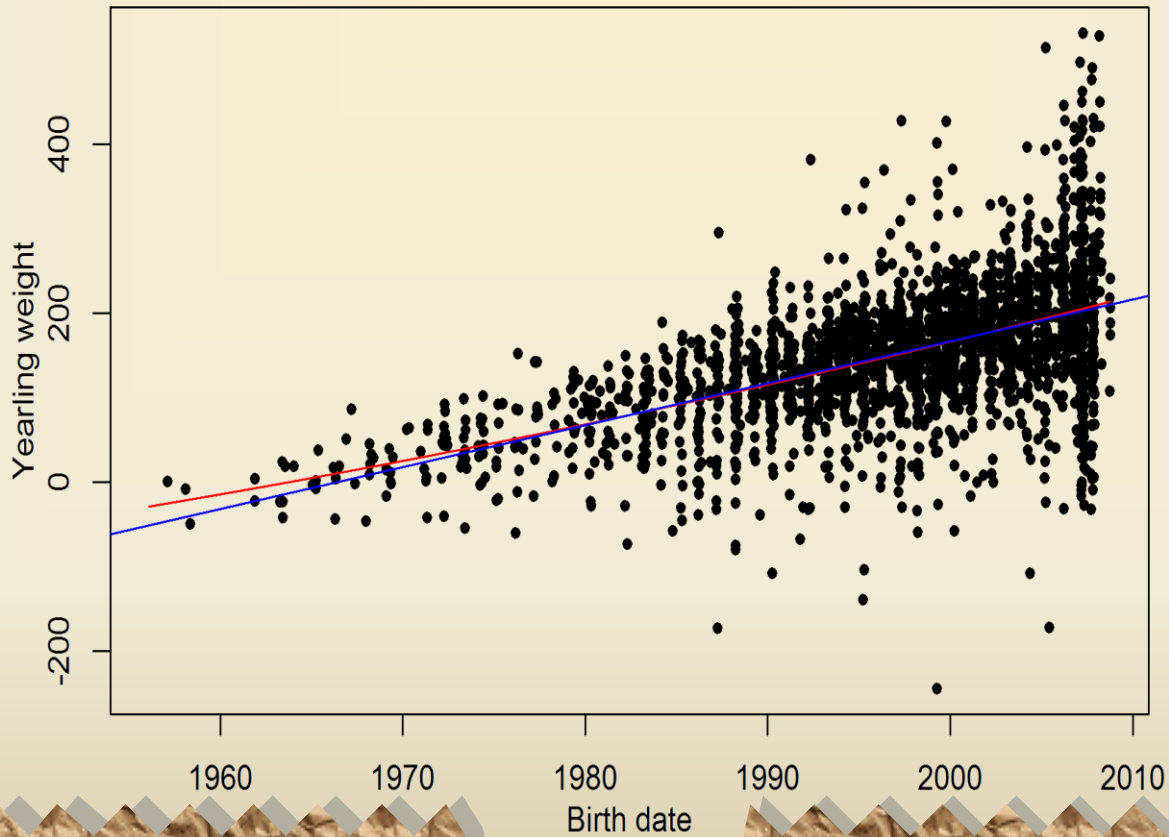
MM Rolf  
Oklahoma State University

BP Kinghorn  
University of New England, NSW, Australia

MD MacNeil  
Miles City, MT

Deregressed EBVs for Yearling Weight of 2,755 registered Angus bulls demonstrates that breeders have achieved an average increase of 4.96 lb per year (blue line) over a 50 year period.

Dramatic  
Genetic  
Change

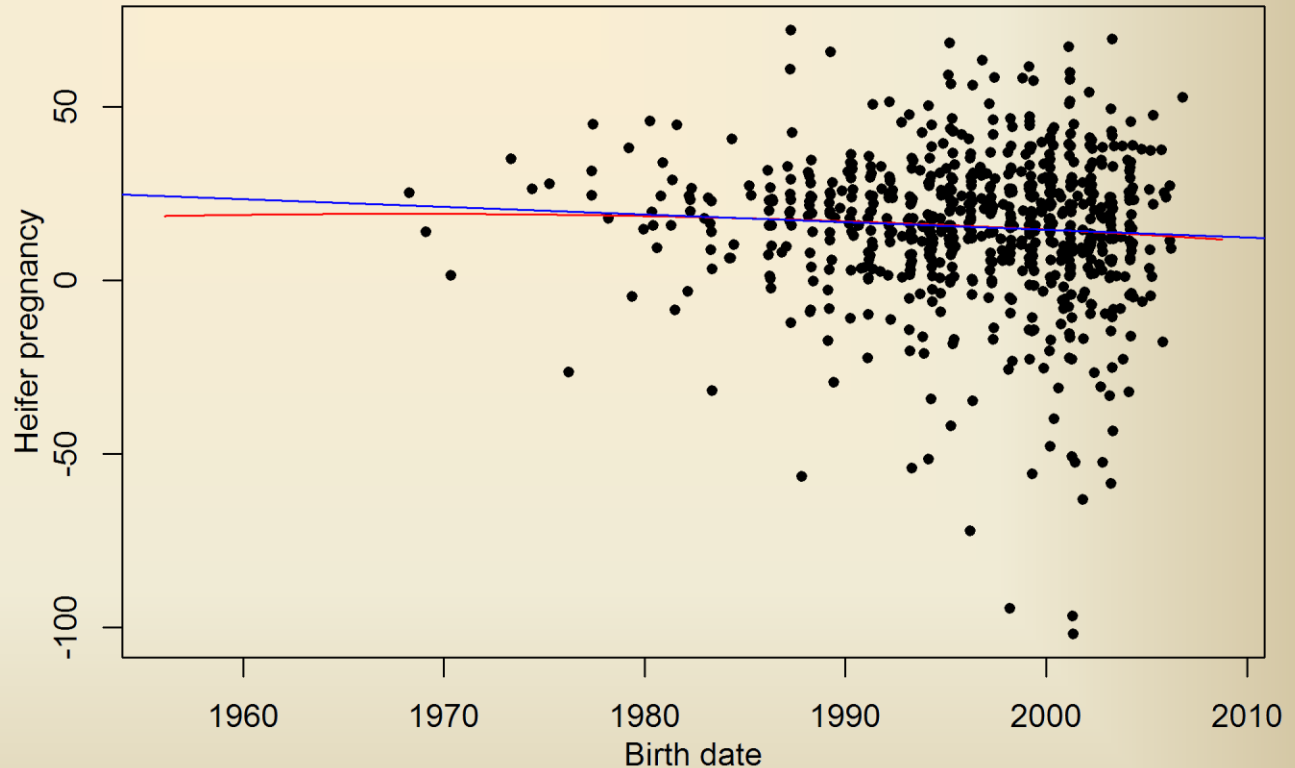


Deregressed EBVs for Heifer Pregnancy Rate for 698 registered Angus bulls indicates that Angus female fertility has decreased by 0.22% per year for about the last 25 years.

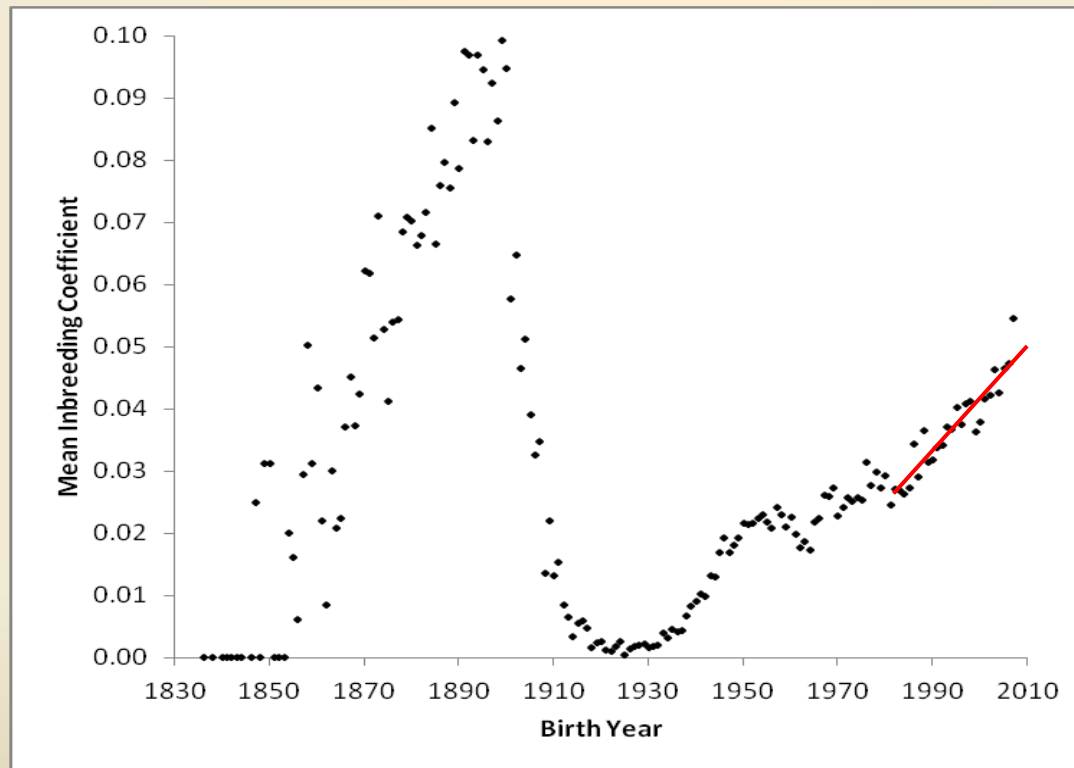
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Possible Cause(s)  
Correlated response  
to selection?

Accumulation of  
inbreeding?



Mean inbreeding coefficients by birth year for 76,083 Angus animals forming a 64 generation pedigree including 3,570 genotyped animals. Effective population size ( $N_e = 116.15 \pm 0.04$ ) was estimated for animals born  $\geq 1980$ .



# Consequences of Inbreeding

- Increases probability of alleles being homozygous
  - As with all lowly heritable traits (like fertility), reduces fitness

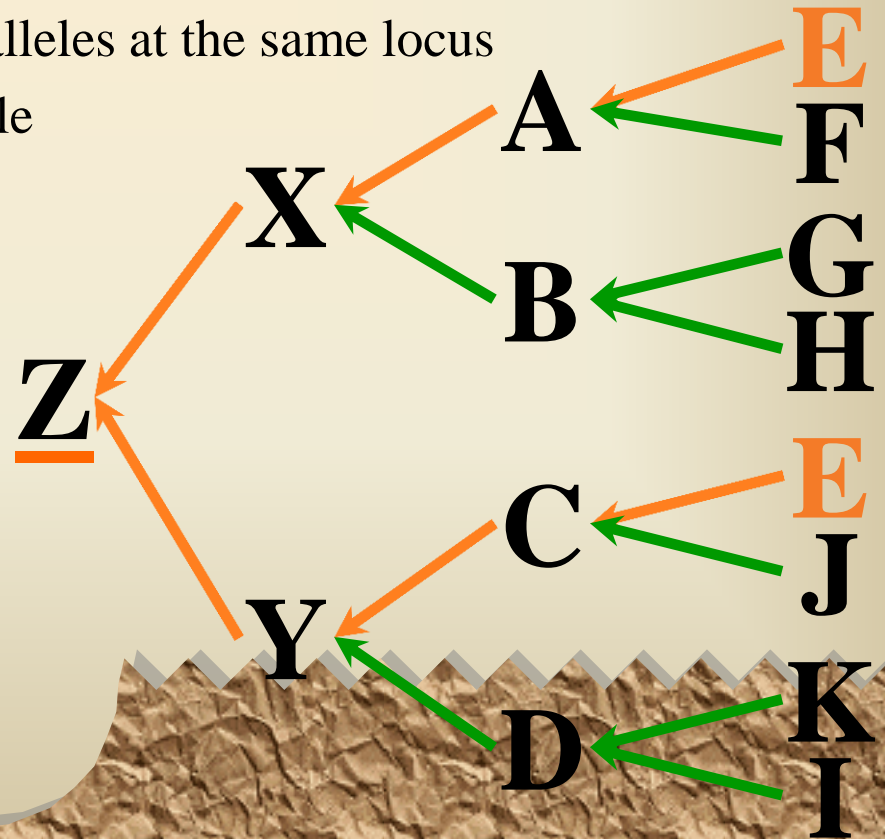
**Table 1.** Summary of heritability and level of heterosis by trait type.<sup>a</sup>

| Trait                    | Heritability | Level of Heterosis   | Inbreeding Depression |
|--------------------------|--------------|----------------------|-----------------------|
| Carcass/end product      | High         | Low<br>(0 to 5%)     | Low                   |
| Skeletal measurements    |              |                      |                       |
| Mature weight            |              |                      |                       |
| Growth rate              | Medium       | Medium<br>(5 to 10%) | Medium                |
| Birth weight             |              |                      |                       |
| Weaning weight           |              |                      |                       |
| Yearling weight          |              |                      |                       |
| Milk production          | Low          | High<br>(10 to 30%)  | High                  |
| Maternal ability         |              |                      |                       |
| Reproduction             |              |                      |                       |
| Health                   |              |                      |                       |
| Cow longevity            |              |                      |                       |
| Overall cow productivity |              |                      |                       |

<sup>a</sup> Adapted from Kress and MacNeil, 1999.

# Effects of Inbreeding Accumulation

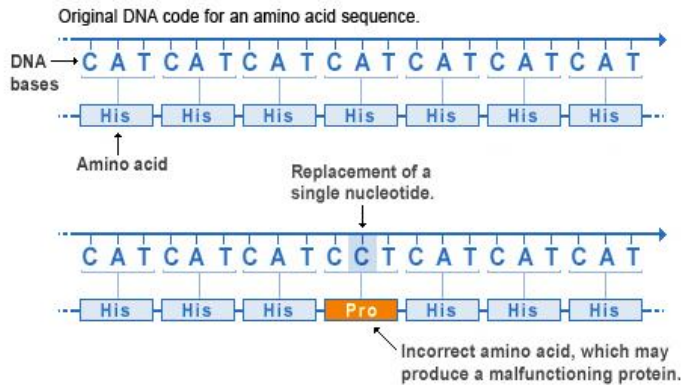
- Increases probability of alleles being homozygous
  - As with all lowly heritable traits, reduces fitness
  - Increases odds of alleles being identical by descent
    - Increases odds of two broken alleles at the same locus
    - Does not cause the broken allele



# Broken Genes

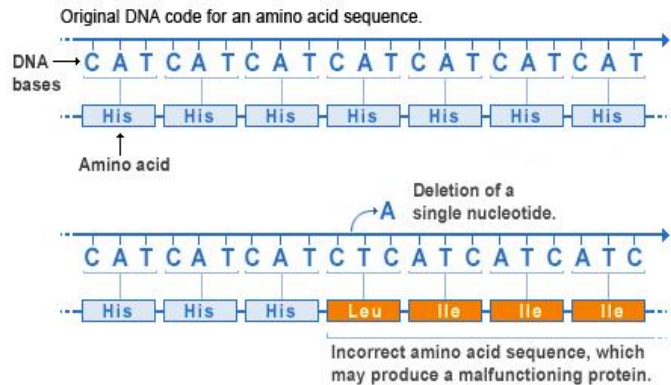
- How do you break a gene?

## Missense mutation



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## Deletion mutation



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### Twenty-One Amino Acids

⊕ Positive ⊖ Negative  
\* Side chain charge at physiological pH 7.4

**A. Amino Acids with Electrically Charged Side Chains**

**Positive**

- Arginine (Arg) **R** (pKa 12.10)
- Histidine (His) **H** (pKa 6.04)
- Lysine (Lys) **K** (pKa 10.87)

**Negative**

- Aspartic Acid (Asp) **D** (pKa 3.71)
- Glutamic Acid (Glu) **E** (pKa 4.15)

**B. Amino Acids with Polar Uncharged Side Chains**

- Serine (Ser) **S** (pKa 9.05)
- Threonine (Thr) **T** (pKa 9.90)
- Asparagine (Asn) **N** (pKa 8.70)
- Glutamine (Gln) **Q** (pKa 9.00)

**C. Special Cases**

- Cysteine (Cys) **C** (pKa 8.14)
- Selenocysteine (Sec) **U** (pKa 10)
- Glycine (Gly) **G** (pKa 9.55)
- Proline (Pro) **P** (pKa 10.47)

**D. Amino Acids with Hydrophobic Side Chain**

- Alanine (Ala) **A** (pKa 9.71)
- Valine (Val) **V** (pKa 9.52)
- Isoleucine (Ile) **I** (pKa 9.60)
- Leucine (Leu) **L** (pKa 9.33)
- Methionine (Met) **M** (pKa 9.28)
- Phenylalanine (Phe) **F** (pKa 9.09)
- Tyrosine (Tyr) **Y** (pKa 9.04)
- Tryptophan (Trp) **W** (pKa 9.34)

frameshifts (not multiple of 3)

Insertion of a single nucleotide.

DNA bases → CATCATCATCATCATCATCAT

Amino acid → His His His **Thr Ser Ser Ser**

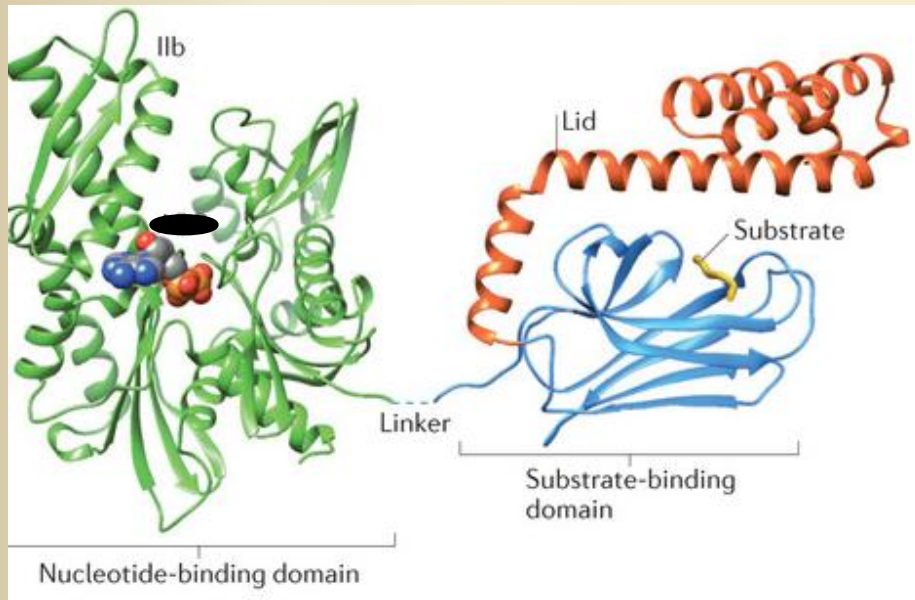
Incorrect amino acid sequence, which may produce a malfunctioning protein.

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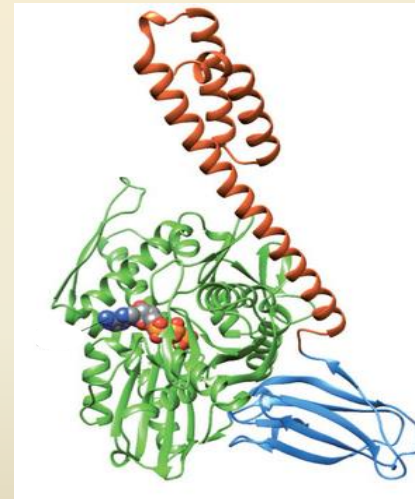


# Results from Broken Genes

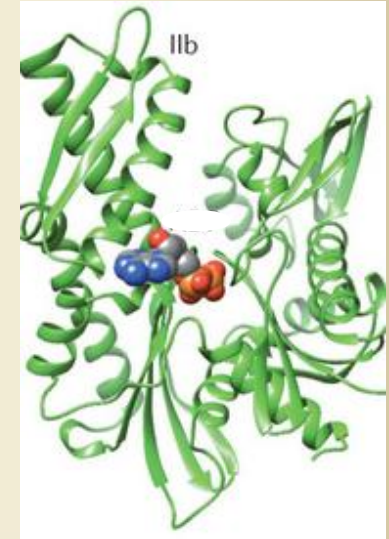
- Truncations, improper folding



Improper Folding



Truncated



# Consequences of Inbreeding

- Called Loss of Function Mutations (LOF)
  - Can be one of two forms
  - Not Critical for Life
    - Will see all genotypes in the population (AA, AB, and BB)
    - Animals may have reduced performance or other deleterious effects, but are functional organisms
  - Critical for Life
    - Animals cannot survive without at least one fully functional version of these genes
    - Presumably calves that are homozygous for a loss of function (LOF) allele will abort, be born dead, die soon after birth, or will never be observed in live animals



# The Case of the Missing Homozygotes

$p$  = frequency of normal allele (0.95)

$q$  = frequency of broken allele (0.05)

$p^2$  = frequency of homozygous normal

$2pq$  = frequency of heterozygotes

$q^2$  = frequency of homozygous broken

Genotype 10,000 animals-

Expect to see:

9,025 homozygous

950 heterozygotes

25 alternate homozygote

Genotype 10,000 animals for lethal-

You would see:

9048 homozygous normal

952 heterozygotes

0 homozygous broken

Need lots of animals to test this!



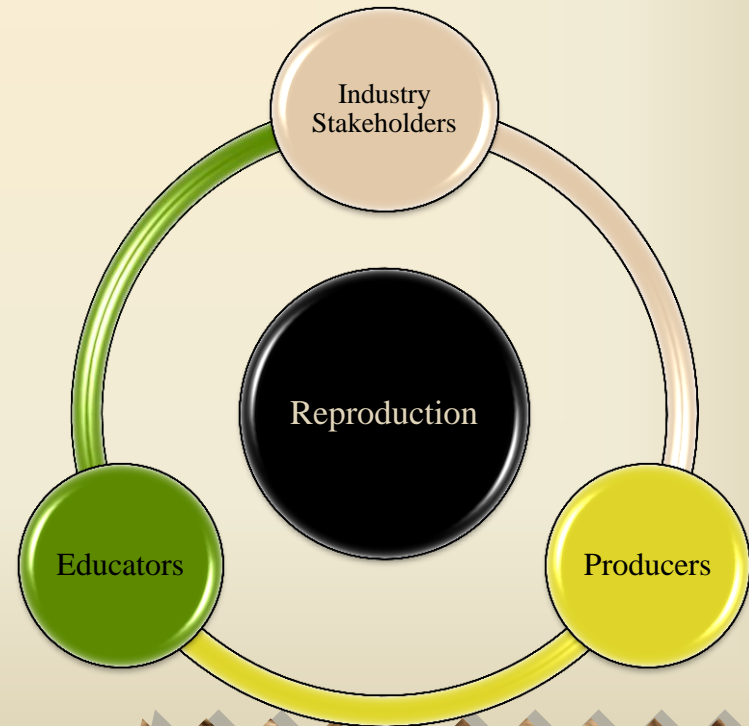
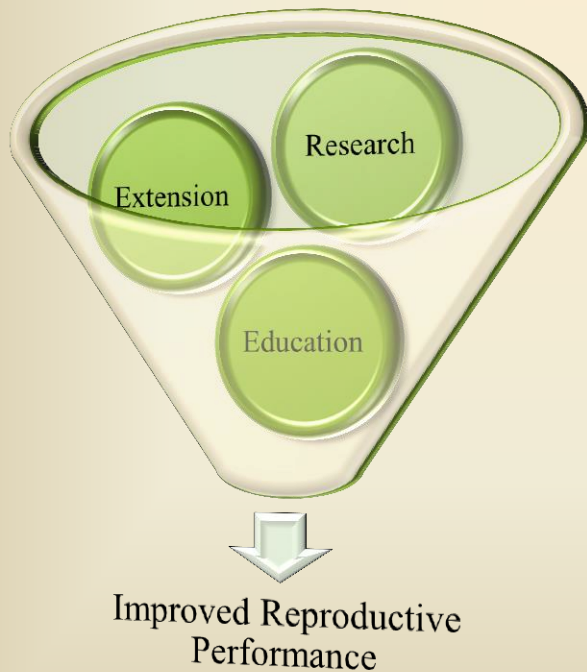
# Project Goals and Background

- Improve reproductive rate of US beef cattle by identifying and managing broken alleles (LOF mutations)
  - Does not sacrifice performance in other ERT
  - Improves overall profitability of the cowherd
- No secret that reproduction is a very important trait in the cowherd
- Managing LOF mutations can assist in
  - Maximizing the number of females that conceive early in the breeding season
  - Maintenance of pregnancies that are achieved



# How to Get There?

- Integrated approach
  - Research, Extension and Education components
  - Partnerships with educators, industry stakeholders and producers



# Specific Aims (Research)

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1

- Sequence highly-influential bulls in Angus and other breeds

2

- Identify candidate LOF alleles never observed in homozygous form

3

- Validate candidate LOF alleles through genotyping large population of phenotyped heifers, remove those found as homozygous

4

- Develop EPD and index selection tools for fertility
- Mate selection software (MateSel)

# Progress to Date

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- Whole Genome Sequencing (bulls)
  - 109 Angus bulls have been sequenced
  - Several other breeds sequenced through cooperative agreements
  - Genomes contributed from collaborators at Genome Canada, USDA/BARC and 18 breed associations
- *In total 267 bulls representing 18 breeds have been sequenced*

# Sequencing Status

| Breed        | N          | Number of reads        | Total bases               | Total coverage | Average coverage |
|--------------|------------|------------------------|---------------------------|----------------|------------------|
| Angus        | 109        | 77,930,820,090         | 7,694,958,893,355         | 2,653.43       | 26.80            |
| Red Angus    | 14         | 4,430,950,144          | 441,846,880,499           | 152.36         | 10.88            |
| Hereford     | 18         | 14,775,544,682         | 1,390,024,023,122         | 479.32         | 26.63            |
| Limousin     | 12         | 3,704,169,818          | 357,264,463,240           | 123.19         | 10.27            |
| Charolais    | 11         | 8,061,833,430          | 802,164,255,493           | 276.61         | 25.15            |
| Simmental    | 11         | 8,902,705,282          | 885,698,817,042           | 305.41         | 27.76            |
| Gelbvieh     | 8          | 6,366,906,096          | 633,479,558,830           | 218.44         | 27.31            |
| Maine Anjou  | 5          | 4,061,220,172          | 403,867,224,031           | 139.26         | 27.85            |
| Romagnola    | 4          | 901,554,762            | 89,666,842,589            | 30.92          | 7.73             |
| Shorthorn    | 2          | 1,370,128,728          | 136,274,291,678           | 46.99          | 23.50            |
| Beefmaster   | 10         | 8,351,392,646          | 830,865,082,100           | 286.51         | 28.65            |
| Holstein     | 25         | 3,224,948,436          | 320,796,806,908           | 110.62         | 4.42             |
| Jersey       | 9          | 1,399,450,902          | 139,150,036,295           | 47.98          | 5.33             |
| Ndama        | 1          | 739,233,320            | 73,483,493,461            | 25.34          | 25.34            |
| Brahman      | 11         | 1,871,667,422          | 167,772,161,118           | 57.85          | 5.26             |
| Nelore       | 8          | 1,668,006,036          | 165,728,918,125           | 57.15          | 7.14             |
| Gir          | 6          | 1,583,737,248          | 157,449,065,756           | 54.29          | 9.05             |
| Bison        | 3          | 3,242,100,744          | 322,544,004,793           | 111.22         | 37.07            |
| <b>Total</b> | <b>267</b> | <b>152,586,369,958</b> | <b>15,013,034,818,435</b> |                |                  |



# Progress to Date

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- Identify candidate LOF alleles
  - Identify and evaluate mutations not found in homozygous form
  - Identify those with predicted disruptive effects on protein structure (frameshifts, deletions, insertions, premature stop codons, non-synonymous mutations)
- Developed SNP chip with predicted LOF DNA sequence variants (DSV) for validation of LOF mutations
  - Remove any that are found in homozygous state in larger population of healthy animals
  - Significant deviations from HWE should also be a clue (missing homozygotes)

# Chip Data Sources

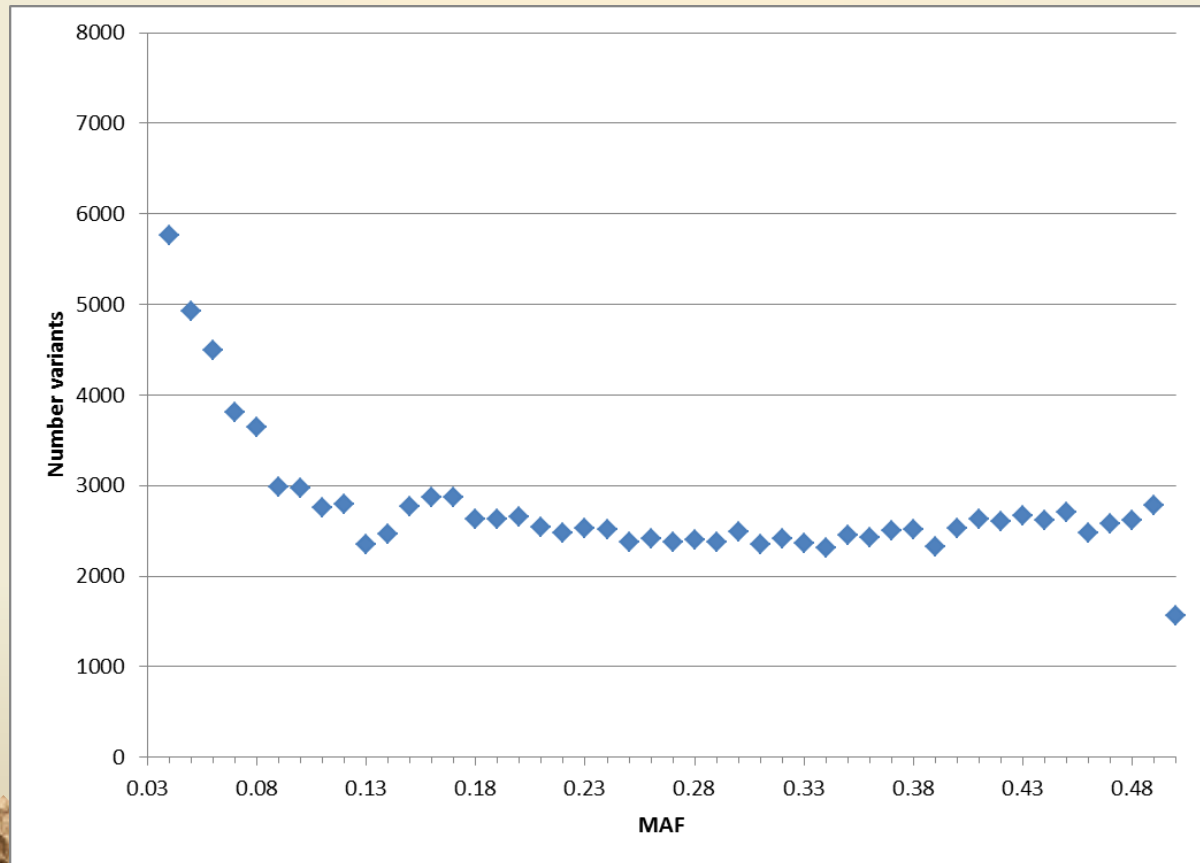
- Chip content for design was from sequences from Mizzou or 1000 bulls project
- Sources of variants:
  - 244 *Bos taurus* genomes
  - 150 *Bos taurus* animals with RNA-Seq data
  - 1000 bulls project variant calls
- Interesting variants classified by type of mutation (non-synonymous, frameshift, premature stop, etc.)
  - “No HOM” classified as variants observed in 2 individuals and no homozygotes observed
    - Within annotation boundaries for gene (not necessarily coding)
    - Variants considered validated if observed in multiple data sources (design pools, any available chips, Affy screening array and dbSNP)
  - Some of these variants not designable (high-freq variants in flanking sequence)
- Detailed information: Jerry Taylor ([taylorjerr@missouri.edu](mailto:taylorjerr@missouri.edu))

# Variant Summary

| Number Variants | Description  |
|-----------------|--|
| 33,730          | Imputation content   |
| 193,503         | Functional content   |
| 227,233         | Total variants   |
|                 |  |
| 31,835          | from HD as part of imputation content                        |
| 22,183          | from SNP50 as part of imputation content (subset of HD)      |
| 6,395           | from HD as part of functional content                        |
| 443             | from SNP50 as part of functional content (subset of HD)      |
| 38,230          | Total HD   |
| 22,626          | Total SNP50 (subset of HD)                                   |
|                 |  |
| 22,298          | NS Sift deleterious  |
| 48,994          | NS Sift tolerated  |
| 49,627          | NS no sift prediction  |
| 120,919         | Total Non-synonymous (NS) AA substitutions                   |
|                 |  |
| 1,265           | Frameshift indels  |
| 585             | In-Frame indels  |
| 20,402          | UTR  |
| 1,573           | Ensembl ncRNA (snoRNA, miRNA, snRNA, rRNA, Mt_tRNA, Mt_rRNA) |
| 4,081           | Conserved non-coding elements                                |
| 6,378           | Splice (not mutually exclusive)                              |

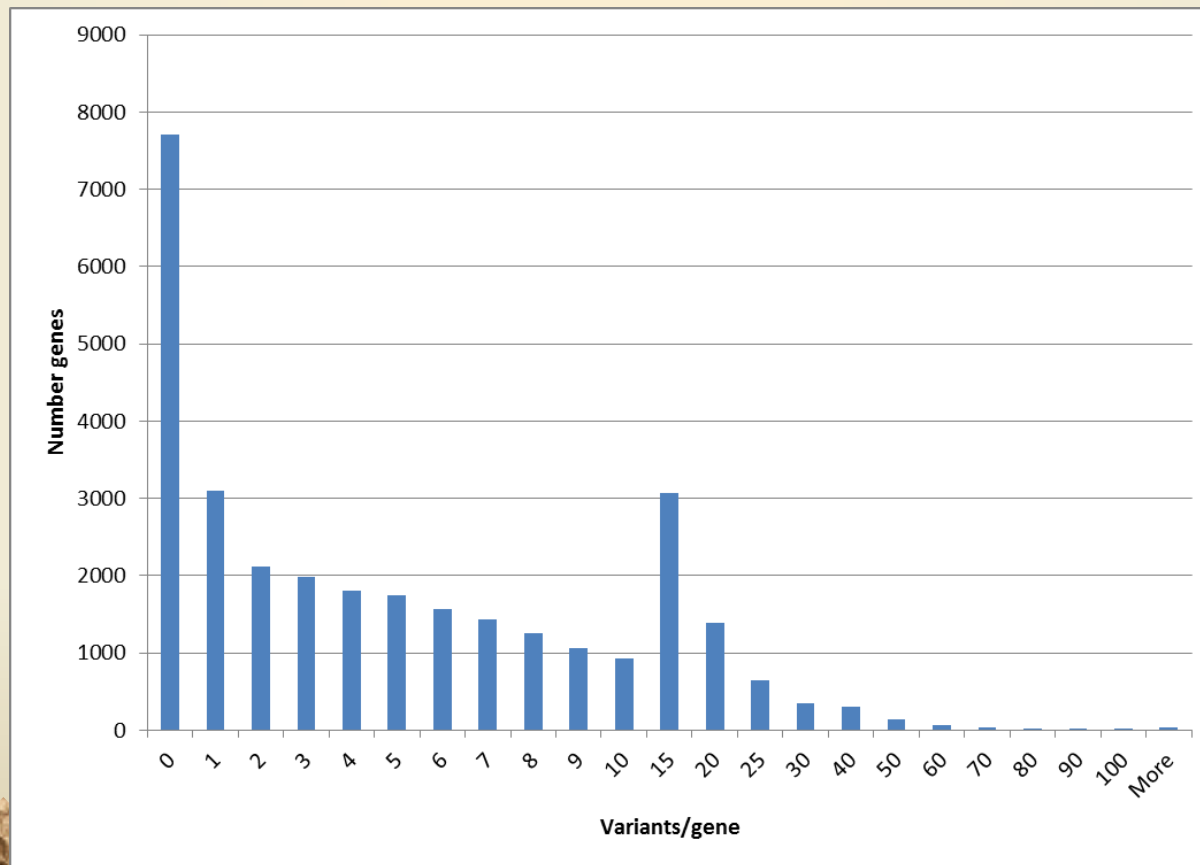
# Distribution of MAF

Biased  
towards  
low  
frequency  
variants



# Number of Genes Represented

Most genes at least 1 variant



|        |                               |
|--------|-------------------------------|
| 23,059 | Genes with at least 1 variant |
|--------|-------------------------------|

|       |                       |
|-------|-----------------------|
| 7,714 | Genes with no variant |
|-------|-----------------------|

# General Design of GGP F250 Assay

- 220K variants (GGP F250)
- 24K for imputation to 50K or greater
- 196K potentially functional variants , biased towards genic regions
- In design now - available to all late Fall 2015

The anticipated cost of the assay will be about \$100 per sample dependent on volume.

Inquiries can be directed to Stewart Bauck at GeneSeek ([SBauck@neogen.com](mailto:SBauck@neogen.com)).

# Progress to Date

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- DNA sampling and phenotypic data collection (heifers)
  - 10,251 Angus heifers have been sampled
    - Complete reproductive data from 44 farms and ranches
  - Genotyping on GGP F250 will be completed in fall 2015



# Progress to Date

- Develop selection indexes that support multi-trait selection, inclusive of fertility traits (MacNeil)
  - Economically rational emphases for fertility, growth, efficiency, and carcass traits upon which to base selection and mating decisions
- Development of decision support software that can incorporate information on LOF alleles and make selection decisions based on a relevant index or breeding objective (Kinghorn)
  - Called MateSel
  - Balance long-term selection gains, inbreeding, and frequency of recessive alleles or carrier to carrier matings
  - MateSel can also show you the ‘opportunity cost’ of imposing non-optimal constraints on mate
    - Selecting against affected progeny more effective than selecting against carriers

Van Eenennaam, A.L., and B. P. Kinghorn. 2014. Use of Mate Selection Software to Manage Lethal Recessive Conditions in Livestock Populations. WCGALP Vancouver, Canada.

[https://asas.org/docs/default-source/wcgalp-posters/408\\_paper\\_9819\\_manuscript\\_1027\\_0.pdf](https://asas.org/docs/default-source/wcgalp-posters/408_paper_9819_manuscript_1027_0.pdf)



# Specific Aims (Extension and Education )

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5

- Develop simulation exercise that demonstrates the effect of DGV for heifer and sire fertility on reproductive performance and profitability (Smith, coming 2016)

6

- Develop a web-based educational training program (Rolf, Smith and Van Eenennaam)

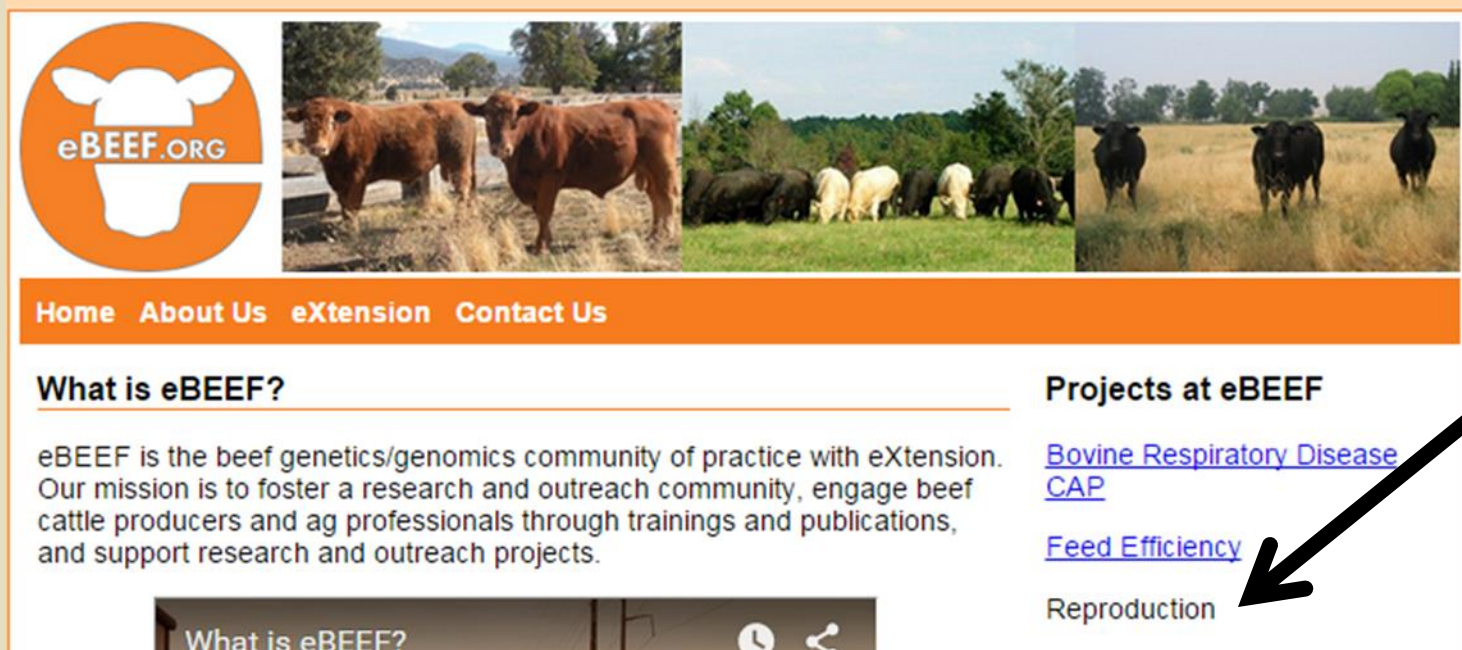
# Educational Modules



|  |  |   |
|--|--|---|
|   | <p>Identification and Management of Alleles<br/>Impairing Heifer Fertility While Optimizing<br/>Genetic Gain in Cattle</p> |  <p>Fact<br/>Sheet</p>   |
|  |    |  <p>Meet Our<br/>Partners</p>  |
| <p><b>Coming Soon.</b></p>   |  |   |



# Educational Resource Links



The screenshot displays the eBEEF.org website interface. At the top left is the eBEEF.ORG logo, which is a white silhouette of a cow's head inside an orange circle. To the right of the logo are three photographs of cattle: two brown cows in a field, a group of black and white cows grazing, and three black cows in a field. Below the images is an orange navigation bar with the links: Home, About Us, eXtension, and Contact Us. The main content area is divided into two columns. The left column is titled "What is eBEEF?" and contains a paragraph of text. The right column is titled "Projects at eBEEF" and lists three categories: "Bovine Respiratory Disease CAP", "Feed Efficiency", and "Reproduction". A large black arrow points from the right side of the page towards the "Feed Efficiency" link. At the bottom of the screenshot, a video player is partially visible with the title "What is eBEEF?".

**eBEEF.ORG**

Home About Us eXtension Contact Us

### What is eBEEF?

eBEEF is the beef genetics/genomics community of practice with eXtension. Our mission is to foster a research and outreach community, engage beef cattle producers and ag professionals through trainings and publications, and support research and outreach projects.

### Projects at eBEEF

- [Bovine Respiratory Disease CAP](#)
- [Feed Efficiency](#)
- Reproduction

What is eBEEF?



This project was supported by Agriculture and Food Research Initiative Competitive Grant no. Agriculture.2013-68004-20364 from the USDA National Institute of Food and Agriculture



United States Department of Agriculture  
National Institute of Food and Agriculture